

## Two New Carbazole Alkaloids from *Murraya koenigii*

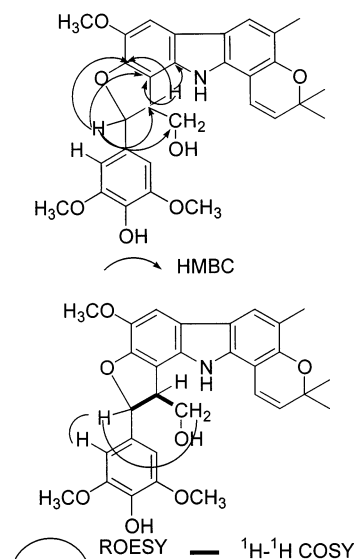
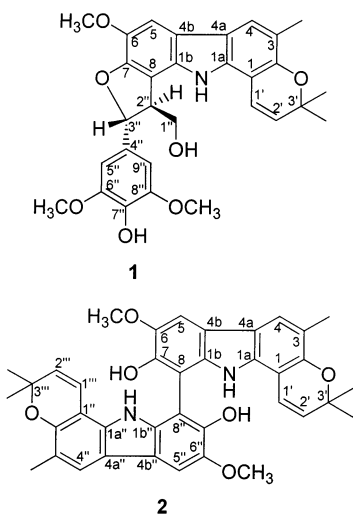
Yun-Song Wang,<sup>†</sup> Hong-Ping He,<sup>†</sup> Yue-Mao Shen,<sup>†</sup> Xin Hong,<sup>†</sup> and Xiao-Jiang Hao<sup>\*,†,‡</sup>

The State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming, Yunnan 650204, People's Republic of China, and Key Laboratory of Chemistry for Natural Products of Guizhou Province and Chinese Academy of Sciences, Guiyang, Guizhou 550002, People's Republic of China

Received October 4, 2002

Two new carbazole alkaloids named murrayanine (**1**) and 8,8''-biskoenigine (**2**) were isolated from *Murraya koenigii*. The structure elucidations for **1** and **2** were carried out on the basis of 1D and 2D NMR experiments. Compound **1** was a novel carbazole alkaloid with a rare phenylpropanyl substitution. Compound **2** was a symmetrical dimer of the carbazole alkaloid koenigine and showed antiosteoporotic activity in the CAT-B model with IC<sub>50</sub> 1.3 μg/mL. The synthesis of **2** from koenigine was carried out through oxidative coupling using a solid state reaction.

Plants of the genus *Murraya* (Rutaceae) are shrubs mainly distributed in Southern Asia.<sup>1</sup> The leaves and bark are used as folk medicines for analgesia and local anesthesia and for the treatment of eczema, rheumatism, and dropsy.<sup>2</sup> The ethanol extract of *M. koenigii* displayed cytotoxic activity against cultured KB cells.<sup>3</sup> In our investigations of the active principles of *M. koenigii*, two novel carbazole alkaloids named murrayanine (**1**) and 8,8''-biskoenigine (**2**) were isolated.



**Figure 1.** Selected <sup>1</sup>H–<sup>1</sup>H COSY, HMBC, and ROESY correlations for compound **1**.

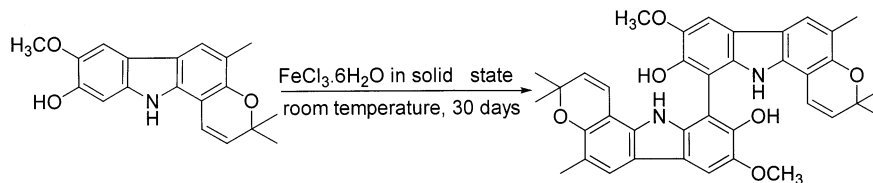
presence of a koenigine moiety<sup>1,4,8–13</sup> and an 11-carbon residue. The latter was confirmed by the EIMS fragment ion at *m/z* 210 (*M* – 307) and the NMR data. The <sup>1</sup>H–<sup>1</sup>H COSY spectrum indicated the presence of the fragment –OCH<sub>2</sub>–CHR<sub>1</sub>–CHR<sub>2</sub>–O–, which was supported by HMBC correlations between H-3'' and C-2'', C-1'' (Figure 1). The appearance of an isolated two-spin system at δ<sub>H</sub> 6.81 (2H, d, *J* = 1.9 Hz) indicated a symmetrically substituted aromatic residue.<sup>14–16</sup> Methoxyl substitutions at C-6'' and C-8'' were revealed by analysis of NMR data, while the C-7'' hydroxyl was determined with the aid of EIMS fragmentation and NMR data.<sup>17–19</sup>

The obvious difference between the NMR data of **1** and koenigine<sup>1,4,8–13</sup> was at C-8; the methine (δ<sub>C</sub> 97.9, d, δ<sub>H</sub> 5.72, 1H, s) in koenigine was replaced by a quaternary carbon (δ<sub>C</sub> 110.9s), indicating that **1** had carbon substitution at C-8.<sup>7,9,12,13</sup> This correlation was supported by <sup>1</sup>H–<sup>13</sup>C long-range correlations between the proton at δ 3.88 (H-2'') and the carbons at δ 148.3 (C-7), 110.9 (C-8), and 131.8 (C-1b), and between the methylene protons at δ 4.02 (2H, d, *J* = 6.0 Hz, H-1'') and the carbon at δ 110.9 (C-8). The connection between C-7 and C-3'' via an oxygen was determined by the <sup>1</sup>H–<sup>13</sup>C long-range correlation between δ<sub>H</sub> 5.46 (H-3'') and the carbon at δ 148.3 (C-7) (Figure 1).

Compounds **1** and **2** give negative reactions with Dragendorff's alkaloid reagent, but produced blue-violet spots with concentrated sulfuric acid on silica gel TLC, which is a characteristic of carbazole alkaloids.<sup>4,5</sup>

Compound **1** was obtained as a brown powder. The HREIMS gave the [M]<sup>+</sup> peak at *m/z* 517.2142, corresponding to the molecular formula C<sub>30</sub>H<sub>31</sub>NO<sub>7</sub>. The IR spectrum showed absorption at ν<sub>max</sub> 3440 and 3410 cm<sup>-1</sup>, indicating the presence of –OH and –NH groups,<sup>6</sup> which was confirmed by the presence of two broad singlets at δ 4.75 and 9.73 in the <sup>1</sup>H NMR spectrum.<sup>7</sup> Inspection of the NMR data (<sup>1</sup>H, <sup>13</sup>C, DEPT, HMQC, and HMBC) indicated the

\* To whom correspondence should be addressed. Tel: 86-871-5219684. Fax: 86-871-5150227. E-mail: xjhao@hotmail.com, yshen@public.km.yn.cn.  
<sup>†</sup> Kunming Institute of Botany.  
<sup>‡</sup> Key Laboratory of Chemistry for Natural Products of Guizhou Province and Chinese Academy of Sciences.



**Figure 2.** Synthesis of **2**.

The relative configurations of C-2'' and C-3'' were determined to be *trans* on the basis of the coupling constants ( $J = 7.8$  Hz) of H-3'' with H-2'' and the ROESY correlations between the protons at  $\delta$  4.02 (H-1'') and 5.46 (H-3'') (Figure 1).<sup>20–24</sup> Thus, the structure of **1**, named murrayanine, was elucidated to be as shown.

Compound **2** was isolated as a brown gum. The HREIMS gave the  $[M]^+$  peak at  $m/z$  616.2585, corresponding to the molecular formula  $C_{38}H_{36}N_2O_6$ . The  $^1H$  and  $^{13}C$  NMR signals were half of that expected on the basis of HREIMS data, suggesting that **2** was a completely symmetrical dimer.<sup>9</sup> The EIMS data suggested that koenigine<sup>4</sup> was the monomer of **2**. The UV and  $^1H$  and  $^{13}C$  NMR spectra of **2** were similar to those of koenigine,<sup>1,4,8–13</sup> indicating that **2** was a dimer of koenigine.<sup>4,7,9</sup> The NMR signals assigned to the C-8 methine ( $\delta_H$  7.43, s,  $\delta_C$  97.9, d) in koenigine were replaced by  $\delta$  105.0 s in **2**, revealing the linkage between C-8 and C-8'', which was further supported by upfield shifts of 2.0 and 0.2 ppm observed for C-7 and C-8a, respectively. Thus, compound **2**, named 8,8''-biskoenigine,<sup>25</sup> was elucidated to be as shown.

To confirm the assigned structure, **2** was synthesized from koenigine through an oxidative coupling reaction, with  $FeCl_3$ , in the solid state (Figure 2).<sup>26–28</sup> This reaction provided a new method for the synthesis of binary carbazole alkaloids for bioactivity research.

Carbazole alkaloids **2** and koenigine were tested using antibacterial (enolpyruvyl transferase, EPT), antifungal (*Candida albicans*, *C. glabrata*, *Aspergillus fumigatus*, YNG), anticancer (cell division cycle 25, CDC25), anti-thrombus (plasminogen activator inhibitor, PAI), and anti-osteoporosis (cathepsin B, CAT-B, caronic anhydrase II, CA-II) bioassays.<sup>29</sup> Compound **2** showed antiosteoporotic activity in the CAT-B model with a  $IC_{50}$  of 1.3  $\mu g/mL$ .

## Experimental Section

**General Experimental Procedures.** IR spectra were measured on a Perkin-Elmer-577 spectrophotometer. MS were performed on an Autospec-3000 spectrometer at 70 eV. The NMR spectra were recorded on Bruker AM-400 and DRX-500 spectrometers.

**Plant Material.** The aerial part of *M. koenigii* was collected in Xishuangbanna, Yunnan Province, P. R. China, in April 1999 and identified by Prof. D. D. Tao of Kunming Institute of Botany. A voucher specimen (No. H98041705) is deposited in the Kunming Institute of Botany, Kunming, China.

**Extraction and Isolation.** The air-dried plant materials (2.5 kg) were extracted with 95% EtOH at ambient temperature for two weeks. The EtOH extract was partitioned between  $H_2O$  and  $CHCl_3$ . An aliquot (100 g) of the  $CHCl_3$ -soluble portion was subjected to repeated column chromatography over silica gel and eluted with a gradient of petroleum ether–EtOAc from 9:1 to 4:6 (v/v) to afford 10 fractions. Fraction 6 (10.2 g) was further subjected to repeated column chromatography over silica gel (eluted with  $CHCl_3$ –EtOAc, 9:1, 8:2, 7:3, 6:4, v/v) and Sephadex LH-20 (eluted with MeOH) to afford koenigine (560 mg) and **2** (52 mg). Fraction 8 (1.2 g) was further purified by column chromatography over silica gel eluted with  $CHCl_3$ – $Me_2CO$  (85:15, v/v) and Sephadex LH-20 eluted with MeOH to yield **1** (7.5 mg).

**Compound (1):** brown powder;  $[\alpha]_D^{25} +8.0$  ( $c$  0.74,  $CH_3OH$ ); UV ( $CH_3OH$ )  $\lambda_{max}$  206, 225, 238, 300, 342 nm; IR (KBr)  $\nu_{max}$  3440, 3410, 2927, 2853, 1733, 999, 907  $cm^{-1}$ ;  $^1H$  NMR ( $CD_3COCD_3$ , 500 MHz)  $\delta$  9.71 (1H, s, NH), 7.59 (1H, s, H-4), 7.47 (1H, s, H-5), 6.81 (1H, d,  $J = 1.9$  Hz, H-9'', 5''), 6.77 (1H, d,  $J = 9.6$  Hz, H-1'), 5.72 (1H, d,  $J = 9.6$  Hz, H-2'), 5.46 (1H, d,  $J = 7.8$  Hz, H-3''), 4.02 (2H, d,  $J = 6.0$  Hz, H-1''), 3.90 (3H, s, OMe-6), 3.88 (1H, dd,  $J = 7.8, 6.0$ , H-2''), 3.80 (6H, s, MeO-8'', 6''), 2.26 (3H, s, Me-3), 1.41 (3H, s, Me-3'), 1.40 (3H, s, Me-3'');  $^{13}C$  NMR ( $CD_3COCD_3$ , 100 MHz)  $\delta$  148.7 (s, C-2), 148.3 (s, C-7), 148.2 (s, C-8'', 6''), 140.2 (s, C-6), 136.2 (s, C-7''), 135.5 (s, C-1a), 132.5 (s, C-4''), 131.8 (s, C-1b), 129.2 (d, C-2'), 120.3 (d, C-4), 118.0 (d, C-1'), 117.8 (s, C-4b), 117.5 (s, C-3), 117.2 (s, C-4a), 110.9 (s, C-8), 104.9 (d C-1), 104.2 (d, C-9'', 5''), 103.7 (d, C-5), 87.7 (d, C-3''), 75.7 (s, C-3'), 63.8 (t, C-1''), 56.6 (q, OMe-6), 56.1 (q, OMe-8'', 6''), 54.0 (d, C-2''), 27.3 (q, Me-3'), 27.1 (q, Me-3'), 15.6 (q, Me-3); EIMS  $m/z$  (%) 517 ( $M^+$ , 100), 502 (10), 499 (15), 484 (25), 418 (30), 396 (17), 368 (10), 210 (10); HREIMS  $m/z$  517.2142 (calcd for  $C_{30}H_{31}NO_7$ , 517.2100).

**Compound (2):**<sup>25</sup> brown gum;  $[\alpha]_D^{25} +139.6$  ( $c$  1.00,  $CHCl_3$ ); UV ( $CH_3OH$ )  $\lambda_{max}$  225, 301, 343 nm; IR (KBr)  $\nu_{max}$  3387, 2934, 2875, 1671, 1458, 1102, 1076, 999, 907  $cm^{-1}$ ;  $^1H$  NMR ( $CD_3COCD_3$ , 500 MHz)  $\delta$  9.56 (1H, s, NH), 7.66 (1H, s, H-4), 7.61 (1H, s, H-5), 6.74 (1H, d,  $J = 9.8$  Hz, H-1'), 5.56 (1H, d,  $J = 9.8$  Hz, H-2'), 4.05 (3H, s, OMe-6), 2.31 (3H, s, Me-3), 1.41 (3H, s, Me-3'), 1.40 (3H, s, Me-3'');  $^{13}C$  NMR ( $CD_3COCD_3$ , 100 MHz)  $\delta$  149.0 (s, C-2), 144.7 (s, C-7), 143.5 (s, C-6), 136.1 (s, C-1a), 135.9 (s, C-1b), 129.2 (d, C-2'), 120.5 (d, C-4), 119.2 (d, C-1'), 118.8 (s, C-4b), 117.5 (s, C-3), 115.4 (s, C-4a), 105.7 (d, C-1), 105.0 (s, C-8), 102.3 (d, C-5), 76.0 (s, C-3'), 57.1 (q, MeO-6), 27.9 (q, Me-3'), 27.2 (q, Me-3'), 16.3 (q, Me-3); EIMS  $m/z$  (%) 616 ( $M^+$ , 100), 601 (56), 308 (10), 293 (36); HREIMS  $m/z$  616.2585 (calcd for  $C_{38}H_{36}N_2O_6$ , 616.2573).

**Synthesis of Compound 2.** A mixture of koenigine (60.9 mg, 0.1 mmol) and  $FeCl_3 \cdot 6H_2O$  (reagent I, 270 mg, 1 mmol) was finely pulverized by agate mortar and pestle and placed in a test tube and kept at room temperature for 30 days. The reaction mixture was quenched with diluted HCl (10 mL) and extracted with  $CHCl_3$  (7 mL, three times). The  $CHCl_3$  extract was subjected to column chromatography over silica gel to produce **2** in 30% yield. The product was identical to natural compound **2** in all respects (Figure 2).

**Acknowledgment.** This work was financially supported by the National Foundation for Outstanding Young Scientists (H.X.J.) (No. 39525025). All spectra were recorded by the analytical group of the Laboratory of Phytochemistry, Kunming Institute of Botany, Chinese Academy of Sciences, and the cooperative laboratory for screening between Kunming Institute of Botany, the Chinese Academy of Sciences, and Bayer AG., Germany, for bioactivity tests.

## References and Notes

- Furukawa, H.; Wu, T. S.; Ohta, T.; Kuoh, C. S. *Chem. Pharm. Bull.* **1985**, *33*, 4132–4138.
- Wu, T. S.; Wang, M. L.; Wu, P. L. *Phytochemistry* **1996**, *43*, 785–789.
- Fiebig, M.; Pezzuto, J. M.; Soejarto, D. D.; Kinghorn, A. D. *Phytochemistry* **1985**, *24*, 3041–3043.
- Bowen, I. H.; Perera, K. P. W. C. *Phytochemistry* **1982**, *21*, 433–437.
- Reisch, J.; Goj, O.; Wickramasinghe, A.; Herath, H. M. T. B.; Henkel, G. *Phytochemistry* **1992**, *31*, 2877–2879.
- Narasimhan, N. S.; Paradkar, M. V.; Chitguppi, V. P.; Kelkar, S. L. *Indian J. Chem.* **1975**, *13*, 993–999.
- Ito, C.; Ohta, H.; Tan, H. T. W.; Furukawa, H. *Chem. Pharm. Bull.* **1996**, *44*, 2231–2235.
- Joshi, B. S.; Kamat, V. N.; Gawad, D. H. *Tetrahedron* **1970**, *26*, 1475–1482.

- (9) Wu, T. S.; Wang, M. L.; Lai, J. S.; Ito, C.; Furukawa, H. *Phytochemistry* **1991**, *30*, 1052–1054.
- (10) Wu, T. S. *Phytochemistry* **1991**, *30*, 1048–1051.
- (11) Furukawa, H.; Wu, T. S.; Kuoh, C. S. *Heterocycles* **1985**, *23*, 1391–1393.
- (12) Ito, C.; Thoyama, Y.; Omura, M.; Kajiura, I.; Furukawa, H. *Chem. Pharm. Bull.* **1993**, *41*, 2096–2100.
- (13) Mukherjee, M.; Mukherjee, S.; Shaw, A. K.; Ganguly, S. N. *Phytochemistry* **1983**, *22*, 2328–2329.
- (14) Martins, R. C. C.; Latorre, L. R.; Sartorelli, P.; Kato, M. J. *Phytochemistry* **2000**, *55*, 843–846.
- (15) Enriquez, R. G.; Chávez, M. A.; Jáuregui, F. *Phytochemistry* **1980**, *19*, 2024–2025.
- (16) Li, S. Y.; Fuchino, H.; Kawahara, N.; Sekita, S.; Satake, M. *J. Nat. Prod.* **2002**, *65*, 262–266.
- (17) Tofern, B.; Jenett-Siems, K.; Siems, K.; Jakupovic, J.; Eich, E. *Phytochemistry* **2000**, *53*, 119–128.
- (18) Zhang, Z. Z.; Guo, D.; Li, C. L.; Zheng, J. H.; Koike, K.; Jia, Z. H.; Nikaido, T. *Phytochemistry* **1999**, *51*, 469–472.
- (19) Yoshikawa, K.; Kimura, Y. K. E.; Arihara, S. *Phytochemistry* **1998**, *49*, 2057–2060.
- (20) Takeda, Y.; Mima, C.; Masuda, T.; Hirata, E.; Takushi, A.; Otsuka, H. *Phytochemistry* **1998**, *49*, 2137–2139.
- (21) Otsuka, H.; Kashima, N.; Nakamoto, K. *Phytochemistry* **1996**, *42*, 1435–1438.
- (22) Apers, S.; Paper, D.; Bürgermeister, J.; Baronikova, S.; Dyck, S. V.; Lemiére, G.; Vlietinck, A.; Pieters, L. *J. Nat. Prod.* **2002**, *65*, 718–720.
- (23) Ullah, N.; Ahmad, S.; Anis, E.; Mohammad, P.; Rabnawaz, H.; Malik, A. *Phytochemistry* **1999**, *50*, 147–149.
- (24) Góngora, L.; Máñez, S.; Giner, R. M.; Recio, M. C.; Gray, A. I.; Ríos, J. L. *Phytochemistry* **2002**, *59*, 857–860.
- (25) Wang, Y. S.; He, H. P.; Hong, X.; Zhao, Q.; Hao, X. J. *Chin. Chem. Lett.* **2002**, *13*, 849–850.
- (26) Lin, G. Q.; Zhang, A. M. *Tetrahedron* **2000**, *56*, 7163–7171.
- (27) Toda, F.; Tanaka, K.; Iwata, S. J. *J. Org. Chem.* **1989**, *54*, 3007–3009.
- (28) Bringmann, G.; Tasler, S.; Endress, H.; Kraus, J.; Messer, K.; Wohlfarth, M.; Lobin, W. *J. Am. Chem. Soc.* **2001**, *123*, 2703–2711.
- (29) Jia, R. R. Transfer, Establishment Screening of Antiosteoporotic Activity Bioassay CAT-B and Anticancer Activity Bioassay CDC25. M.Sc. Thesis, Kunming Institute of Botany, Chinese Academy of Sciences: Kunming, 2002, pp 6–19.

NP020468A